Universida<sub>de</sub>Vigo

# Control of the structure of marine picoplankton communities by turbulence and nutrient supply dynamics



Grupo de

Oceanografía Biolóxica

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# Introduction

#### The importance of phytoplankton



**MODIS Science Team** 





INTRODUCTION

#### Phytoplankton and the biological carbon pump

 $H_2PO_4 + 16NO_3^- + 106CO_2 + 122H_2O \leftrightarrow (C_{106}H_{263}O_{110}N_{16}P_1) + 138O_2$ 





INTRODUCTION

#### **Relevance of nitrogen and supply mechanisms**



#### **Relevance of nitrogen and supply mechanisms**



#### **Turbulence effects over biological data**



**Biological spatial scales** 

Physical spatial scales

Modified from Prairie et al. (2012)

#### How is turbulence measured?



Microstructure shear sensor

#### CTD

#### **Microstructure turbulence profiler (MSS)**



Microstructure shear sensor

Disipation rate of turbulent kinetic energy (E).

CTD

Brunt–Väisälä frequency (N).

#### **Microstructure turbulence profiler (MSS)**



Microstructure shear sensor

Disipation rate of turbulent kinetic energy (E).

CTD

Brunt–Väisälä frequency (N).

Vertical diffusivity (K<sub>z</sub>):  

$$Kz = 0.2 \frac{\varepsilon}{N^2}$$

Osborn (1980)

#### Nutrient stock and nutrient flux



#### Nutrient stock and nutrient flux in Atlantic Ocean



The variability in nutrient stock **can be disconnected** from changes in nutrient supply (Mouriño-Carballido *et al.* 2011)

#### **Competition dynamics**



INTRODUCTION

Villamaña et al., 2019

# Why picoplankton?

- The most abundant organisms in the ocean
- Picophytoplankton often dominate primary production in gyres
- Expected future expansion of gyres area in a future ocean scenario



# Which groups do picoplankton include ?



<b>Bacterioplankton</b>	<u>Cyanobacteria</u>	<b><u>Picoeukaryotes</u></b>
LNA	Prochorococcus	
HNA	Synechococcus	

# Environmental control factors in the distribution of picophytoplankton



# Environmental control factors in the distribution of picophytoplankton

Temperature



Light

BACKGROUND

#### **Environmental control factors in the** distribution of picophytoplankton

Temperature



Temperature & Light are the main control factors of the regional distributions of both *Prochlorococcus* and *Synechococcus* (Flombaum *et al.*, 2013).

Light

#### Environmental control factors in the distribution and activity of picoplankton



Reproduced from Mouriño-Carballido et al. (2016)

#### **Biogeochemical implications of picoplankton**

- Aggregation (Richardson & Jackson, 2007):
  - Avaliable for copepods (fastsinking fecal pellets).
  - Increase sinking velocity
  - Southern ocean (Lomas & Moran, 2011):
    - Pico and nanoplankton export 33±27% of the total carbon



Guidi et al, 2016



# Hypothesis and objectives

### Hypothesis

Nutrient supply dynamics (constant versus variable supply) controls the structure of marine picoplankton communities.

### **Objectives**

- 1. To **quantify** the role of **temperature**, **light**, and **nitrate fluxes** as factors controlling the distribution of autotrophic and heterotrophic picoplankton subgroups.
- To describe the ecological niches of the various components of the picoplankton community.
  - . To explore the effect of nitrate supply dynamics on the competitive dynamics of two model marine picophytoplankton species, namely, the cyanobacterium *Synechococcus* sp. and the picoeukaryote *Micromonas pusilla*.
- 4. To build a prediction model and obtain the first **climatology** of **nitrate diffusion** into the **euphotic zone**.
- 5. To **predict** the change in the structure of **picophytoplankton communities** (the cyanobacteria to picoeukaryotes ratio) in a **future ocean scenario**.

#### **Research approach**





**Chapter II:** Factors controlling picoplankton community structure

#### **Objectives**

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#### **Dataset of biological & physical data (2006-2015)**



- ❑ Vertical dissipation rate (Kz)
- Nutrients
- □ PAR (Satellite)
- Picoplankton biomass (Cytometry)

#### Nitrate diffusive flux



#### Nitrate diffusive flux



#### Nitrate advective flux



- MATERIAL & METHODS

CHAPTER II

#### Nitrate advective flux



# Analysis

### Analysis

#### Generalized Additive Models (GAM)

 $yj = I + s(SST) + s(PAR) + s(log(NO_3Flux)) + Error$
# Analysis

### Generalized Additive Models (GAM)

 $yj = I + s(SST) + s(PAR) + s(log(NO_3Flux)) + Error$ 



### Variability in NO<sub>3</sub> flux, control factors and biomass



### **Relevance of control factors in biomass** groups (GAM)



### **Relevance of control factors in biomass** groups (GAM)



### Niche partitioning



### Niche partitioning





**Chapter III:** *Micromonas pusilla* and *Synechococcus* competition under constant and dynamic conditions

### Dominance of picoplankton groups vs mixing and NO<sub>3</sub>



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### **Steady state and chemostats**



### **Competition experiments**



Population monitoring was carried using Flow Cytometry

### **Experimental design**

Groups

□ Synechococcus (RCC-2366)

□ Micromonas pusilla (RCC-450)



Yearly average surface chlorophyll-a

# **Experimental design**

#### Groups

Synechococcus (RCC-2366)

□ Micromonas pusilla (RCC-450)

- Fully-aclimated populations
  - □ Modified PCRS-11 medium (N:P, 5-1)
  - $\Box$  Light: 100  $\mu$ E
  - ☐ Temperature: 21°C
  - □ Steady-state (Dilution rate: 0.2 d<sup>-1</sup>)

### Perturbation(5 µM NO<sub>3</sub>)

- $\Box$  0.5 pulses d<sup>-1</sup>
- $\Box$  1 pulses d<sup>-1</sup>\*
- $\Box$  2 pulses d<sup>-1</sup>
- $\Box$  3 pulses d<sup>-1</sup>



Yearly average surface chlorophyll-a



Sartorius Biostat Plus

### **Uptake experiments**

Similar light and temperature conditions
Short NO<sub>3</sub> incubations (Bulk concentration)
[NO<sub>3</sub>]: 0.5, 1, 1.5, 2.5, 5, 10, 25 μM.
Gentle filtration (Ø 0.45 μm)



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### **Ecological modelling and calibration**



- Droop model
- Delayed Rejection Adaptative Metropolis Algorithm (DRAM):
  - Uptake and batch experiments parameters used as initial parameters.
  - Use one experiment to calibrate and use the other 3 to test.

### **Competition experiments – Time series**



### **Competitive exclusion rate**



### **Modelling - Calibration**





### **Modelling - Calibration**



### **Modelling - Calibration**



### **Competition experiments - Modelling**





**Chapter IV:** Climatology of the vertical nutrient supply and future cyanobacteria to picoeukaryotes ratio

# **Objectives**

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### **Dataset of microstructure turbulence (2006-2015)**



#### 16 cruises; 181 stations

- □ 181 Microturbulence (MST, 0-300 m)
- □ Nitrate concentration (0-200 m):
  - □ 172 Observations
  - □ 6 WOA09 database
  - □ 3 Nitrate-density relationship

# Multivariable fractional polynomial method (MFP)

### **Independent variables**

Stratification	Nitrate	Chlorophyll-a
SST	sNO <sub>3</sub>	DCM
SSS	nitraD	maxChl-a
MLD	grNO <sub>3</sub>	sChl-a
$maxN^2$		
dmaxN <sup>2</sup>		
avrN <sup>2</sup>		

### **MFP** algorithm



### **Future scenario (2100)**



Lewandowska et al., 2014

### **Future scenario (2100)**



### **Future scenario (2100)**



# Variability in NO<sub>3</sub> gradient, K, NO<sub>3</sub> flux and sChl-*a*



### **Collinearity in the dataset**



### **Collinearity in the dataset**



### **Collinearity in the dataset**



### Multivariable fractional polynomial method (MFP)

	R <sup>2</sup> -adj	AIC
Tropical and subtropical		
FNO <sub>3</sub> = f( <b>grNO<sub>3</sub></b> , SSS, sNO <sub>3</sub> , avrN <sub>2</sub> )	0.75	143
FNO <sub>3</sub> = f( <b>grNO<sub>3</sub>, SST</b> )	0.41	189
NW Mediterranean		
$FNO_3 = f(avrN_2)$	0.68	72
FNO <sub>3</sub> = f( <b>SST, sChla</b> )	0.64	77
NW Galician upwelling		
FNO <sub>3</sub> = f( <b>grNO<sub>3</sub>,</b> maxChla)	0.64	77
FNO <sub>3</sub> = f( <b>grNO</b> 3)	0.51	110
Antartic		
<i>FNO</i> <sub>3</sub> = <i>f</i> ( <b>SST</b> )	0.75	38
Global		
FNO <sub>3</sub> = f( <b>SST, grNO3, sChla</b> , DCM)	0.55	545
FNO₃= f( <b>SST, grNO3, sChla</b> )	0.52	553

### Multivariable fractional polynomial method (MFP)

	R <sup>2</sup> -adj	AIC	
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## Prediction of NO<sub>3</sub> turbulent diffusion



SST from WOA13

grNO<sub>3</sub> from WOA13

sChla from Globecolour (1998-2017)

## Prediction of NO<sub>3</sub> turbulent diffusion + observations

Log<sub>10</sub> Flux NO<sub>3</sub>



## Prediction of NO<sub>3</sub> diffusion for $40^{\circ}N - 40^{\circ}S$

Log<sub>10</sub> Flux NO<sub>3</sub>

 $NO_3$  Flux < 1 mmol m<sup>-2</sup> d<sup>-1</sup> sChl-a < 1 mg m<sup>-3</sup>



- **RESULTS** 

CHAPTER IV

# Relevance of diffusive nitrogen fluxes in tropical and subtropical areas





<sup>1</sup>This study

# Relevance of diffusive nitrogen fluxes in tropical and subtropical areas





<sup>1</sup>This study <sup>2</sup>Fernández-Castro et al. (2015)



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20% ratio phyto respiration to GP (Geider, 1992)
23% DOC production (Teira et al, 2001)
Variable stoichiometry (Galbraith & Martiny, 2015)



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### Present and future of cyanoB/pEuk ratio







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2100









#### **OBJECTIVE I**

To quantify the role of temperature, light, and nitrate fluxes as factors controlling the

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#### **CONCLUSION I**

**Temperature** and **nitrate supply** were **more relevant than light** in predicting the biomass of most picoplankton subgroups, except for *Prochlorococcus* and low-nucleic-acid (LNA) prokaryotes, for which irradiance also played a significant role.

#### **OBJECTIVE II**

To describe the ecological niches of the various components of the picoplankton community.

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#### **CONCLUSION II y III**

*Prochlorococcus* and LNA prokaryotes were more abundant in warmer waters where the nitrate fluxes were low, *Synechococcus* and high-nucleic-acid (HNA) bacteria prevailed in cooler environments characterized by intermediate or high levels of nitrate supply, and finally the niche of picoeukaryotes was defined by low temperatures and high nitrate supply.

Nitrate supply was the only factor that allowed the distinction among the ecological niches of all autotrophic and heterotrophic picoplankton subgroups.

#### **OBJECTIVE III**

To explore the **effect** of **nitrate supply dynamics** on the competitive dynamics of two model marine picophytoplankton species, namely, the cyanobacterium *Synechococcus* sp. and the picoeukaryote *Micromonas pusilla*.

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#### **CONCLUSION IV, V y VI**

**Nitrate supply dynamics controlled the outcome of competition** between the cyanobacterium *Synechococcus* and the picoeukaryote *M. pusilla*.

Under continuous nitrate limitation conditions (steady-state), *M. pusilla* was outcompeted by *Synechococcus sp.*, the result of the competition was reversed in nutrient supply dynamics scenarios.

The rate of competitive exclusion of *Synechococcus* was a linear function of the frequency of nitrate pulses, demonstrating that there is a window of opportunity for the coexistence of both species.

#### **OBJECTIVE IV**

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#### **CONCLUSION VII y VIII**

A model including **three predictors** (surface temperature, nitrate vertical gradient, and surface chlorophyll-*a*) **explained 57%** of the **variance** in the nitrate diffusive flux.

Average nitrate diffusion for oligotrophic regions between  $40^{\circ}$ N- $40^{\circ}$ S (~20 Tmol N y<sup>-1</sup>) was comparable to the sum of global estimates of nitrogen fixation, fluvial fluxes and atmospheric deposition.

#### **OBJECTIVE V**

To **predict** the change in the structure of **picophytoplankton communities** (the cyanobacteria to picoeukaryotes ratio) in a **future** global change **scenario**.

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#### CONCLUSION IX

The predicted **decrease of nitrate supply** in tropical and subtropical areas as the result of global change

(~20%), would produce an increase in the cyanobacteria to picoeukaryotes biomass ratio of 8%.

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## THANK YOU FOR YOUR ATTENTION





